

The Effect of Exogenous Adenosine on Functional Injury Caused by Hydrogen Peroxide in the Isolated Rat Heart

GURO VALEN and *JARLE VAAGE

Department of Surgery, University of Tromsø, Tromsø, Norway

Accepted by Professor Dr H. Esterbauer

(Received November 24th, 1994; in revised form, May 19th, 1995)

Adenosine is an endogenous cardioprotective substance. The present study examines whether exogenous adenosine attenuates cardiac injury induced by oxidative stress. Rat hearts (Langendorff model) were perfused with H_2O_2 (180 μM) for 10 min, then recovered for 60 min ($n = 10$). In other groups adenosine 55 μM , 110 μM , or 220 μM ($n = 10$ in each) was given in addition to H_2O_2 throughout perfusion. Control perfusion with Krebs Henseleit only ($n = 7$), adenosine 110 μM throughout perfusion ($n = 7$), and adenosine 110 μM as an intervention ($n = 7$) was performed. The hearts were paced at 320 beats/min. Left ventricular systolic (LVSP) and end-diastolic (LVEDP) pressures were measured together with coronary flow (CF), and left ventricular developed pressure (LVDP = LVSP - LVEDP) was calculated. H_2O_2 decreased LVSP from 105 ± 8 to 60 ± 5 mmHg (mean \pm SEM) after 10 min infusion ($p < 0.008$). Adenosine did not attenuate the decrease of LVSP. LVEDP increased from 0 to 59 ± 10 mmHg ($p < 0.004$) and 62 ± 11 mmHg 5 and 15 min after end of infusion of H_2O_2 , respectively. Neither 55 μM nor 220 μM adenosine inhibited the H_2O_2 -induced increase of LVEDP. Adenosine 110 μM attenuated the increase after 15 (15 ± 4 mmHg, $p < 0.004$) and 25 min observation (26 ± 7 mmHg, $p < 0.012$). Adenosine did not attenuate the reduction of LVDP. CF initially increased during infusion of H_2O_2 , thereafter decreased. Hearts given

adenosine had higher basal CF, and CF did not increase after H_2O_2 . Control perfusion with adenosine, given throughout perfusion or as an intervention, increased CF and tended to increase LVSP. In summary, adenosine did not inhibit H_2O_2 -induced depression of contractility or reduction of CF. One concentration of adenosine (110 μM) attenuated H_2O_2 -induced impairment of relaxation. Exogenous adenosine does not have an important influence on functional injury caused by exogenous oxidants.

Key words: Adenosine, antioxidant, hydrogen peroxide, ischaemia-reperfusion, toxic oxygen metabolites

INTRODUCTION

Ischaemia-reperfusion of the heart induces an inflammatory response with a cascade of injurious mediators, of which toxic oxygen metabolites (TOM) may be the most important.¹⁻⁵ Cardioprotective substances, such as adenosine, are also released from the ischaemic-reperfused heart.^{6,7} Adenosine attenuates myocardial

Correspondence to and *current address: Guro Valen, Department of Thoracic Surgery, Karolinska Hospital, S-17176 Stockholm, Sweden, FAX: + 46-8-322701 TEL: + 46-8-7292000

stunning,⁸ is suggested to initiate and mediate preconditioning,⁷ reduces infarct size,⁷ and inhibits microvascular stunning.⁸⁻¹⁰ Adenosine has several mechanisms of action: Induces coronary vasodilation, reduces myocardial oxygen demand, increases uptake of glucose, and replenishes depleted ATP-stores.^{8,11,12} Furthermore, adenosine protects the endothelium by inhibiting platelet activation, leukocyte adherence and extravasation, and inhibits formation and release of TOM from activated leukocytes.^{8, 12-15} Adenosine has also been suggested to inhibit the cardiotoxic effects of TOM by adenosine A₁ receptor activation.¹⁶

At present adenosine receptor agonists are not clinically available. Adenosine levels in patients can be increased by infusion,¹⁷ or by pharmacologically increasing the contents of adenosine in ischaemic tissue.¹⁸ Consequently, it is important to investigate if adenosine directly attenuates oxidant injury.

This study investigates the effects of adenosine on cardiac dysfunction induced by exogenous TOM in isolated rat hearts. Hearts were perfused with H₂O₂, and three different concentrations of adenosine were added to the perfusate.

METHODS

Heart Perfusion

The study was approved of by the Ethical Committee for Animal Research at the University of Tromsø, Tromsø, Norway. Male Wistar rats (250–300 g) were anaesthetized with diethyl ether. Heparin (200 IU) was injected into the femoral vein. The hearts were excised through a median sternotomy and placed in ice-cold buffer during preparation for aortic cannulation. The average time of ischaemia before start of perfusion was 80 seconds. Modified Langendorff perfusion was performed with Krebs Henseleit buffer (NaCl 118.5 mM, NaHCO₃ 25.0 mM, KCl 4.7 mM, KH₂PO₄ 1.2 mM, MgSO₄ · 7H₂O 1.2 mM, Glucose

· H₂O 11.1 mM, CaCl₂ · 2H₂O 2.4 mM). Perfusion pressure was kept constant at 100 cm H₂O, and the perfusate was continuously gassed with 5% CO₂ and 95% O₂. Heated water jackets around the perfusate reservoirs and heart chamber maintained the temperature at 37°C. Two perfusate reservoirs were employed to rapidly change perfusates at intervention. Isovolumetric recordings of left ventricular peak systolic (LVSP) and end-diastolic (LVEDP) pressures were made via a balloon in the left ventricle, introduced through the left atrium. The volume of the balloon (approximately 0.1 ml H₂O) was adjusted in the height of the left ventricle before insertion into each heart to obtain a pressure of 0 mmHg. Hearts that did not have LVEDP of 0 mmHg after 15 min stabilization were discarded. Left ventricular developed pressure (LVDP = LVSP – LVEDP) was calculated. Coronary flow (CF) was measured by timed collections of the coronary effluent. The hearts were paced via an electrode in the left ventricular epicardium at a frequency of 320 beats/min to overcome the negative chronotropic effects of adenosine. Adenosine (crystalline, Sigma Chemical Company, MO, USA) was dissolved in buffer and administered via one perfusate reservoir throughout perfusion unless otherwise stated. H₂O₂ (3% solution) was added to buffer alone or buffer with adenosine and administered via the second perfusate reservoir.

Experimental Protocol

After start of perfusion the hearts were stabilized for 20 min before intervention (defined as time 0). Only hearts with LVSP between 60–160 mmHg, LVEDP 0 mmHg and CF over 6 ml/min after 15 min stabilization were included in one of the following groups:

- 1) (n = 10) H₂O₂ (180 μM) was given from time 0 for 10 min, followed by a 60 min recovery period.
- 2) (n = 10) H₂O₂ given as described above, but with adenosine (55 μM) added to the buffer throughout perfusion.

- 3) (n = 10) H₂O₂ as in group 1, with addition of adenosine 110 µM.
- 4) (n = 10) H₂O₂ as in group 1, with adenosine 220 µM.
- 5) (n = 7) Control perfusion for 70 min with buffer only.
- 6) (n = 7) Control perfusion with adenosine 110 µM throughout perfusion.
- 7) (n = 7) Control perfusion with adenosine 110 µM added from time 0 and onwards.

Measurements of LVSP, LVEDP, LVEDP, and CF were made after 15 min stabilization (time -5), at time 0, and after 5, 10, 15, 25, 40, and 70 min observation. Only values after 0, 10, 25, and 70 min are presented in the tables for didactic reasons. Some of the hearts perfused with H₂O₂ had severe arrhythmias (asystolia or ventricular fibrillation) during recovery. Severe arrhythmias were evaluated from the pressure curve as a pressure generation <4 mmHg. Values from hearts with severe arrhythmias were excluded from the haemodynamic comparisons. The number of hearts being compared at each time point is thus n = 10 - number of arrhythmic hearts at that time shown in Table 1.

Statistics

A Mann-Whitney test for comparison of independent samples was used to evaluate differences between groups, and Wilcoxon's Signed Rank test

TABLE 1 The number of Langendorff-perfused rat hearts with ventricular fibrillation or asystolia after infusion of H₂O₂ (180 µM) for 10 minutes from time 0, followed by recovery for 60 min. The effects of H₂O₂ alone (group 1, n = 10), or H₂O₂ in addition to adenosine 55 µM (group 2, n = 10), adenosine 110 µM (group 3, n = 10), or adenosine 220 µM (group 4, n = 10) are shown.

Time (min)	0	10	25	70
Group 1	0	2	2	4
Group 2	0	0	2	7
Group 3	0	0	1	0
Group 4	0	1	3	3

There were no statistical differences in comparison between group 1 and the other groups.

was used for evaluation within groups at different time points. The p-values were adjusted according to the Bonferroni method,¹⁹ and p<0.0125 was considered significant at a p<0.05 level in comparison between groups 1-4. For comparison between groups 5-7, p<0.017 was considered significant. In evaluation within groups at different time points p<0.008 was considered significant. Values are presented as mean ± SEM.

RESULTS

Arrhythmias

Severe arrhythmias were observed in few hearts during administration of H₂O₂. After return to buffer perfusion, severe arrhythmias which were not modified by adenosine occurred. The number of hearts in groups 1-4 with severe arrhythmias at the various time points are shown in Table 1. There were no significant differences between groups.

Left Ventricular Systolic Pressure

At time 0 LVSP was similar in groups 1-4 (Table 2), and in groups 5-7 (Table 3). H₂O₂ decreased LVSP in group 1 (p<0.008 after 10 min), and LVSP improved during recovery (Table 2). Addition of adenosine (55 µM, 110 µM, or 220 µM, groups 2-4) did not attenuate the H₂O₂-induced depression of LVSP (Table 2).

There was a gradual reduction of LVSP during perfusion in group 5 (Table 3). Control perfusion with adenosine (group 6) tended to increase LVSP compared to group 5, significantly so after 25 min (p<0.016) (Table 3). When adenosine was given as an intervention (group 7), LVSP did not increase (Table 3).

Left Ventricular End-diastolic Pressure

LVEDP was 0 mmHg in all groups at time 0. H₂O₂ increased LVEDP to 28 ± 10 mmHg after 10 min. After end of H₂O₂-perfusion LVEDP continued to

TABLE 2 Left ventricular systolic pressure (LVSP, mmHg), left ventricular developed pressure (LVDP, mmHg) and coronary flow (CF, ml/min) in Langendorff-perfused rat hearts exposed to H₂O₂ for 10 min (group 1), and H₂O₂ with addition of adenosine 55 μ M (group 2), 110 μ M (group 3), or 220 μ M (group 4). n = 10 in each group initially, but values from hearts with severe arrhythmias are excluded (see Table 1 for number).

	0 min	10 min	25 min	70 min
LVSP				
Group 1	105 \pm 8	60 \pm 5*	84 \pm 7	73 \pm 6
Group 2	105 \pm 5	51 \pm 6	93 \pm 7	81 \pm 7
Group 3	104 \pm 4	50 \pm 4	77 \pm 4	68 \pm 5
Group 4	124 \pm 8	73 \pm 5	99 \pm 6	73 \pm 7
LVDP				
Group 1	105 \pm 8	43 \pm 5*	35 \pm 3*	39 \pm 2
Group 2	105 \pm 5	32 \pm 3	42 \pm 6	51 \pm 6
Group 3	104 \pm 4	46 \pm 5	49 \pm 5	43 \pm 5
Group 4	124 \pm 8	25 \pm 4**	41 \pm 9	36 \pm 9
CF				
Group 1	14 \pm 11	15 \pm 1	6 \pm 0.3*	5 \pm 0.5
Group 2	21 \pm 1**	18 \pm 1	8 \pm 1	10 \pm 2
Group 3	17 \pm 1**	16 \pm 1	8 \pm 1	6 \pm 1
Group 4	20 \pm 1**	11 \pm 1**	7 \pm 1	5 \pm 1

Values are mean \pm SEM. * denotes $p < 0.05$ compared to initial value, ** denotes $p < 0.05$ compared to group 1.

increase, to 59 \pm 10 mmHg 5 min after end of H₂O₂ (15 min observation, $p < 0.004$), and to 62 \pm 11 mmHg after 25 min observation (Figure 1). Addition of adenosine 55 μ M and 220 μ M (groups 2 and 4) did not inhibit the H₂O₂-induced increase of LVDP (Figure 1). However, adenosine 110 μ M attenuated the increase of LVDP after 15 (15 \pm 4 mmHg, $p < 0.004$ compared to group 1) and 25 min observation (26 \pm 7 mmHg, $p < 0.012$). There was no difference between groups after 70 min

observation (Figure 1). LVDP remained 0 mmHg throughout perfusion in groups 5–7 (data not shown).

Left Ventricular Developed Pressure

Before start of intervention (time 0) LVDP was equal to LVSP. H₂O₂ decreased LVDP after 10 min infusion ($p < 0.008$), and LVDP was still decreased after 25 min observation ($p < 0.008$)

TABLE 3 Left ventricular systolic pressure (LVSP, mmHg) and coronary flow (CF, ml/min) in Langendorff-perfused rat hearts. The effects of control perfusion with buffer only (group 5, n = 7), with adenosine 110 μ M throughout perfusion (group 6, n = 7), and adenosine 110 μ M given from time 0 (group 7, n = 7) are shown.

	0 min	10 min	25 min	70 min
LVSP				
Group 5	93 \pm 8	88 \pm 7	87 \pm 5	78 \pm 5
Group 6	109 \pm 5	112 \pm 5	106 \pm 4**	83 \pm 4
Group 7	96 \pm 7	115 \pm 7	112 \pm 7	85 \pm 5
CF				
Group 5	12 \pm 1	10 \pm 0.5	9 \pm 0.3	8 \pm 0.4
Group 6	21 \pm 1**	21 \pm 1**	21 \pm 2**	21 \pm 2**
Group 7	12 \pm 1	17 \pm 1**	16 \pm 1**	11 \pm 2

Values are mean \pm SEM. ** denotes $p < 0.05$ compared to group 5.

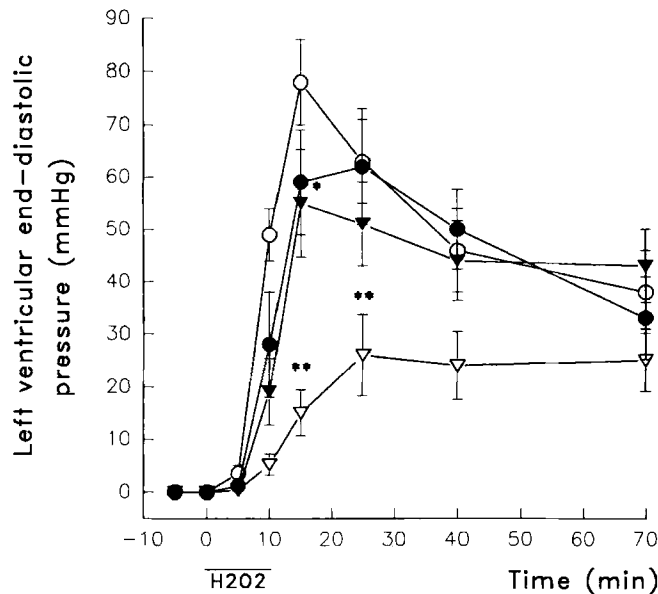


FIGURE 1 Left ventricular end-diastolic pressure in isolated rat hearts perfused with H₂O₂ (180 μM) for 10 min followed by recovery for 60 min (●—●), or H₂O₂ with addition of adenosine 55 μM (▼—▼), 110 μM (▽—▽), or 220 μM (○—○) given throughout perfusion. n = 10 initially in all groups, but values from hearts with severe arrhythmias are excluded (see Table 1 for numbers) * denotes p < 0.05 compared to initial value, ** denotes p < 0.05 compared to H₂O₂ alone.

(Table 2). There was no attenuation of the H₂O₂-induced depression of LVDP in groups 2 and 3 (Table 2). Adenosine 220 μM (group 4) tended to decrease LVDP further, significant after 10 min (p < 0.01) (Table 2).

In control perfused hearts (groups 5–7) LVDP remained the same as LVSP throughout perfusion (Table 3).

Coronary Flow

Adenosine given from start of perfusion increased CF compared to perfusion with Krebs Henseleit only at time 0. CF was higher in group 2 (p < 0.003), group 3 (p < 0.002), and group 4 (p < 0.002) than in group 1 (Table 2). H₂O₂ increased CF to 22 ± 1 ml/min after 5 min infusion (p < 0.004 compared to initial value). CF then decreased compared to initial value after 25 min observation (p < 0.008) (Table 2). CF did not increase in groups 2–4 after infusion of H₂O₂. CF was actually reduced after

10 min of H₂O₂ in group 4 (p < 0.001 compared to group 1) (Table 2).

There was a gradual reduction of CF in group 5 (Table 3). Adenosine 110 μM throughout perfusion (group 6) increased CF compared to controls at time 0 (p < 0.003), and at all other times of observation (p < 0.002 at all time points) (Table 3). Addition of adenosine at time 0 (group 7) increased CF after 5, 10, 15, 25, and 40 min (p < 0.002, p < 0.002, p < 0.002, p < 0.002 and p < 0.016, respectively) (Table 3).

DISCUSSION

To investigate if exogenous adenosine had direct effects on oxidant stress, infusion of H₂O₂ for 10 minutes was employed as a convenient and reproducible model of TOM-induced injury. H₂O₂ has been measured in cardiac tissue after ischaemia-reperfusion injury to isolated rat and rabbit

hearts,^{20,21} the level corresponding to loss of ventricular function during reperfusion.²⁰ H₂O₂ may be the main injurious TOM released by activated leukocytes.^{22,23} Exogenous H₂O₂ dose-dependently influences cardiac function, ranging from only coronary vasodilation to severe impairment of left ventricular function (unpublished results). In the present study a high concentration of H₂O₂ was chosen, in order to detect a possible attenuation of oxidant injury by adenosine.

The exact mechanism of injury in this model is presently unknown. H₂O₂ may have direct toxic effects, or other TOM may be generated. We have previously inhibited all functional impairment of isolated rat hearts caused by H₂O₂ with catalase,²⁴ a scavenger of H₂O₂,¹ and with thiourea,²⁵ a hydroxyl radical scavenger with some effect on H₂O₂.^{26,27} Recently production of the hydroxyl radical was measured in isolated rat hearts perfused with H₂O₂.²⁸ Inhibition of the hydroxyl radical reduced biochemical and ultrastructural injury, while catalase additionally inhibited functional depression.²⁸ It is possible that H₂O₂ is the main injurious oxygen metabolite in the present study.

Three different concentrations of adenosine were given from start of stabilization in order to saturate the hearts, and throughout perfusion in addition to H₂O₂. The concentrations have previously protected against ischaemia-reperfusion injury in buffer-perfused, isolated hearts.^{29,30} The hearts were paced in order to eliminate the inhibitory effects of adenosine on the conduction system.^{8,11} Adenosine did not modify the H₂O₂-induced reductions of LVSP or LVDP. However, 110 µM adenosine attenuated the increase of LVEDP. Adenosine has previously delayed the onset of ischaemic contracture in models of ischaemia-reperfusion injury.^{10,30,31} Our finding is in accordance with this. However, as only one of three relatively close concentrations of adenosine influenced the increase of LVEDP, the interpretation of this finding is difficult. Adenosine may have a bell-shaped dose-response curve in oxidant injury of the heart, explaining why the highest dose (220 µM adenosine) had no beneficial

effect. Indeed the highest dose tended to exacerbate oxidant injury, since it decreased both LVDP and CF after perfusion with H₂O₂ for 10 min.

H₂O₂ increased CF after 5 min infusion. This increase was not seen when adenosine was given in addition to H₂O₂. H₂O₂ increases CF by release of endothelial derived relaxing factor (EDRF),³² while vasodilation induced by adenosine is independent of EDRF.³³ The lack of H₂O₂-induced vasodilation in hearts pretreated with adenosine could be due to the hearts being maximally dilated before intervention, and not by inhibition of the H₂O₂-induced vasodilation per se. Adenosine did not influence reduction of CF during recovery.

Control perfusion with adenosine (110 µM) increased CF and tended to increase LVSP. This is in accordance with well known inotropic and vasodilatory properties of adenosine.^{8,11}

Severe arrhythmias were observed after end of, but not during, infusion of H₂O₂. TOM may cause reperfusion arrhythmias.³⁴ In the present study either other TOM than H₂O₂, or intracellular injury secondary to H₂O₂ (membrane function deficiency, lipid peroxidation, disturbed calcium homeostasis) most likely mediate arrhythmias. Adenosine protects against ischaemia-reperfusion induced arrhythmias in other models.³⁵ There were no differences in occurrence of severe arrhythmias in hearts receiving H₂O₂ alone or in combination with adenosine in the present study.

Few works have attempted to elucidate if adenosine has direct antioxidant effects. A recent work by Karmazyn & Cook,¹⁶ infusing 100 µM H₂O₂ for 30 min in isolated rat hearts with addition of two adenosine A₁ receptor agonists, concluded that adenosine A₁ receptor stimulation reduced functional injury induced by H₂O₂, possible by preservation of high-energy phosphates and adenine nucleotide contents.¹⁶ However, Karmazyn & Cook employed small experimental groups (n = 5 or less) which were evaluated with parametric statistics.¹⁶ A severe model of H₂O₂-induced functional injury with stop of pressure-generating beating was employed.¹⁶ Possibly the effects of

selective adenosine A₁ receptor activation and administration of exogenous adenosine are unlike.

In conclusion, three different concentrations of adenosine did not inhibit the H₂O₂-induced depression of contractility or occurrence of severe arrhythmias. One concentration of adenosine attenuated the impairment of relaxation induced by H₂O₂, but the other two did not. Adenosine did not influence H₂O₂-induced changes in coronary flow. An antagonistic effect against oxidant injury does not appear to be a main component of the cardioprotective actions of adenosine.

Acknowledgements

G.V. has been a research fellow of the Norwegian Research Council. Further financial support was provided by the University of Tromsø.

References

1. P.A. Southorn and G. Powis (1988) Free radicals in medicine. Involvement in human disease. *Mayo Clinic Proceedings*, **63**, 390–406.
2. R. Bolli (1992) Postischemic myocardial "stunning": Pathogenesis, pathophysiology, and clinical relevance. In *Myocardial protection. The pathophysiology of reperfusion injury* (eds D.M. Yellon and R.B. Jennings), Raven Press Ltd., New York, pp. 105–150.
3. J.M. Downey and D.M. Yellon (1992) Do free radicals contribute to myocardial cell death during ischemia-reperfusion? In *Myocardial protection. The pathophysiology of reperfusion injury* (eds D.M. Yellon and R.B. Jennings), Raven Press Ltd., New York, pp. 35–58.
4. J. Vaage and G. Valen (1993) Pathophysiology and mediators of ischemia-reperfusion injury with special reference to cardiac surgery. *Scandinavian Journal of Thoracic and Cardiovascular Surgery*, **Suppl. 41**, 1–18.
5. G. Valen and J. Vaage (1993) Toxic oxygen metabolites and leukocytes in ischemia-reperfusion injury. *Scandinavian Journal of Thoracic and Cardiovascular Surgery*, **Suppl. 41**, 19–29.
6. J. Parrat (1993) Endogenous myocardial protective (antiarrhythmic) substances. *Cardiovascular Research*, **27**, 693–702.
7. J.M. Downey, G.S. Liu and J.D. Thornton (1993) Adenosine and the anti-infarct effects of preconditioning. *Cardiovascular Research*, **27**, 3–8.
8. M. Kitakaze, M. Hori and T. Kamada (1993) Role of adenosine and its interaction with alpha adrenoceptor activity in ischaemic and reperfusion injury of the myocardium. *Cardiovascular Research*, **27**, 18–27.
9. B. Olafsson, M.B. Forman, D.W. Puett, A. Pou, C.U. Cates, G.C. Friesinger and R. Virmani (1987) Reduction of reperfusion injury in the canine preparation by intracoronary adenosine: importance of the endothelium and the no-reflow phenomenon. *Circulation*, **76**, 1135–1145.
10. D.G. Babbitt, R. Virmani and M.B. Forman (1989) Intracoronary adenosine administered after reperfusion limits vascular injury after prolonged ischemia in the canine model. *Circulation*, **80**, 1388–1399.
11. B.B. Fredholm and A. Sollevi (1986) Cardiovascular effects of adenosine. *Clinical Physiology*, **6**, 1–21.
12. R.M. Berne (1993) Adenosine – a cardioprotective and therapeutic agent. *Cardiovascular Research*, **27**, 2.
13. B.N. Cronstein, S.M. Kubersky, G. Weissmann and R. Hirschhorn (1987) Engagement of adenosine receptors inhibits hydrogen peroxide release by activated human neutrophils. *Clinical Immunology and Immunopathology*, **42**, 76–85.
14. M.B. Grisham, L.A. Hernandez and D.N. Granger (1989) Adenosine inhibits ischemia-reperfusion-induced leukocyte adherence and extravasation. *American Journal of Physiology*, **257**, H1334–H1339.
15. M.B. Forman, C.E. Velasco and E.K. Jackson (1993) Adenosine attenuates reperfusion injury following regional myocardial ischaemia. *Cardiovascular Research*, **27**, 9–17.
16. M. Karmazyn and M.A. Cook (1992) Adenosine A₁ receptor activation attenuates cardiac injury produced by hydrogen peroxide. *Circulation Research*, **71**, 1101–1110.
17. A. Öwall, J. Ehrenberg, L.-Å. Brodin, A. Juhlin-Dannfeldt and A. Sollevi (1993) Effects of low-dose adenosine on myocardial performance after coronary artery bypass surgery. *Acta Anaesthesiologica Scandinavica*, **37**, 140–148.
18. S.F. Bolling, M.A. Groh, A.M. Mattson, R.A. Grinage and K.P. Gallagher (1992) Acadesine (AICA-riboside) improves postischemic cardiac recovery. *Annals of Thoracic Surgery*, **54**, 93–98.
19. S. Wallenstein, C.L. Zucker and J.L. Fleiss (1980) Some statistical methods useful in circulation research. *Circulation Research*, **47**, 1–9.
20. J.M. Brown, M.A. Grosso, G.J. Whitman, A. Banerjee, L.S. Terada, J.E. Repine and A.H. Harken (1989) The coincidence of myocardial reperfusion injury and hydrogen peroxide production in the isolated rat heart. *Surgery*, **105**, 496–501.
21. M. Schlafer, K. Brosamer, J.R. Forder, R.H. Simon, P.A. Ward and C.M. Grum (1990) Cerium chloride as a marker of hydrogen peroxide in reperfused ischemic hearts. *Journal of Molecular and Cellular Cardiology*, **22**, 83–97.
22. S.J. Weiss, J. Young, A.F. LoBuglio, A. Slivka and N.F. Nimeth (1981) Role of hydrogen peroxide in neutrophil-mediated destruction of cultured endothelial cells. *Journal of Clinical Investigation*, **68**, 714–721.
23. G.M. Vercelotti, S.P. Severson, P. Duane and C.F. Moldow (1991) Hydrogen peroxide alters signal transduction in human endothelial cells. *Journal of Laboratory and Clinical Medicine*, **117**, 15–24.
24. G. Valen, J. Kaszaki, I. Szabo, S. Nagy and J. Vaage (1993) Toxic oxygen metabolites and ischemia-reperfusion increase histamine synthesis and release in the isolated rat heart. *Journal of Molecular and Cellular Cardiology*, **25**, 31–40.
25. G. Valen, E. Eriksson, B. Risberg and J. Vaage (1993) Reactive oxygen intermediates and ischemia-reperfusion injury release tissue plasminogen activator from isolated rat hearts. *Thrombosis Research*, **76**, 113–121.
26. R.B. Fox (1984) Prevention of granulocyte-mediated oxidant lung injury in rats by a hydroxyl radical scavenger, dimethylthiourea. *Journal of Clinical Investigation*, **74**, 1456–1464.
27. M. Wasil, B. Halliwell, M. Grootveld, C.P. Moorhouse,

- D.C.S. Hutchison and H. Baum (1987) The specificity of thiourea, dimethylthiourea and dimethyl sulphoxide as scavengers of hydroxyl radicals. *Biochemical Journal*, **243**, 867–870.
28. G. Takemura, T. Onodera, R.W. Millard and M. Ashraf (1993) Demonstration of hydroxyl radical and its role in hydrogen-peroxide induced myocardial injury: hydroxyl radical dependent and independent mechanisms. *Free Radicals in Biology and Medicine*, **15**, 13–25.
 29. S.W. Ely, R.M. Mentzer, R.D. Lasley, B.K. Lee and R.M. Berne (1985) Functional and metabolic evidence of enhanced myocardial tolerance to ischemia and reperfusion with adenosine. *Journal of Thoracic and Cardiovascular Surgery*, **90**, 549–556.
 30. S.F. Bolling, L.E. Bies, K.P. Gallagher and E.L. Bove (1989) Enhanced myocardial protection with adenosine. *Annals of Thoracic Surgery*, **47**, 809–815.
 31. R.B. Lasley, J.W. Rhee, D.G.L. Wylen van and R.M. Mentzler (1990) Adenosine A₁ receptor mediated protection of the globally ischemic isolated rat heart. *Journal of Molecular and Cellular Cardiology*, **22**, 39–47.
 32. T. Skjelbakken, G. Valen and J. Vaage (1992) The role of endothelium derived relaxing factor in the cardiac effects of hydrogen peroxide. *Journal of Vascular Research*, **29**, p. 200 (abstract).
 33. J.L. Dinerman and J.L. Mehta (1990) Endothelial, platelet and leukocyte interactions in ischemic heart disease: Insights into potential mechanisms and their clinical relevance. *Journal of American College of Cardiology*, **16**, 207–222.
 34. D.J. Hearse (1992) Myocardial injury during ischemia and reperfusion. In *Myocardial protection. The pathophysiology of reperfusion and reperfusion injury* (eds. D.M. Yellon and R.B. Jennings), Raven Press, New York, pp. 13–33.
 35. C.L. Wainwright and J.R. Parrat (1993) Effects of R-PIA, a selective A₁ adenosine agonist, on haemodynamics and ischaemic arrhythmias in pigs. *Cardiovascular Research*, **27**, 84–89.